

The Coleopterist

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Notes Review

The Coleopterist

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Acupalpus maculatus Schaum, 1860: another carabid new to Britain from Dungeness

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In March 2002, John Paul sent MGT a few carabids to check. Amongst them were three *Acupalpus* specimens from Dungeness, which I compared to the series of *Acupalpus parvulus* (Sturm, 1825) (still known to many British coleopterists as *A. dorsalis* (Fabricius, 1787)) in my collection, and also to a French specimen of *Acupalpus maculatus* Schaum which I collected in June 2000. I was to receive two shocks: first that John's specimens matched *A. maculatus*, and second that I also had three specimens lying unrecognised in my own collection, standing as *A. parvulus* and also collected at Dungeness! Subsequently, several other coleopterists have discovered specimens in their possession, dating back to 2 October 1996. Targeted visits in 2002 have revealed that the species is still present.

IDENTIFICATION

Acupalpus maculatus may be identified using the keys of Jeannel (1941), Freude *et al.* (1976) or Hürka (1996). The identity of British specimens has also been checked by comparison to continental specimens standing over *A. maculatus* in the collections of the Natural History Museum, London.

Elytral pattern

Most of the British specimens seen have a distinctive pattern on the elytra (Fig. 1). Both *A. parvulus* and *A. maculatus* have the apical two-thirds to three-quarters of the elytra dark, with the suture and margins paler yellow-brown. On *A. parvulus* this leaves a conspicuous solidly pale area at the base of the elytra. On *A. maculatus*, this pale area is divided in two by a dark bar that connects the central dark patch to the shoulder (specifically to the base of the sixth elytral interval), isolating a pale spot beside the scutellum from the pale side margin. Jeannel (1941) and Hürka (1996) both make it clear that both species are variable in their colouration and some variation is apparent amongst the specimens seen. Most notably, the palest specimens of *A. maculatus* may have a faint dark mark at the base of the sixth elytral interval, connected to the central dark patch by the merest shadow of a dark bar; without careful scrutiny, such specimens resemble typical *A. parvulus*.

Shape of pronotum

A. maculatus typically shows a less transverse pronotum than *A. parvulus*, being almost as long as broad. Jeannel (1941) and Hürka (1996) both use pronotum shape as an important character in their keys, presumably because it is a more consistent character than the elytral colour pattern. However, British specimens of *A. maculatus*

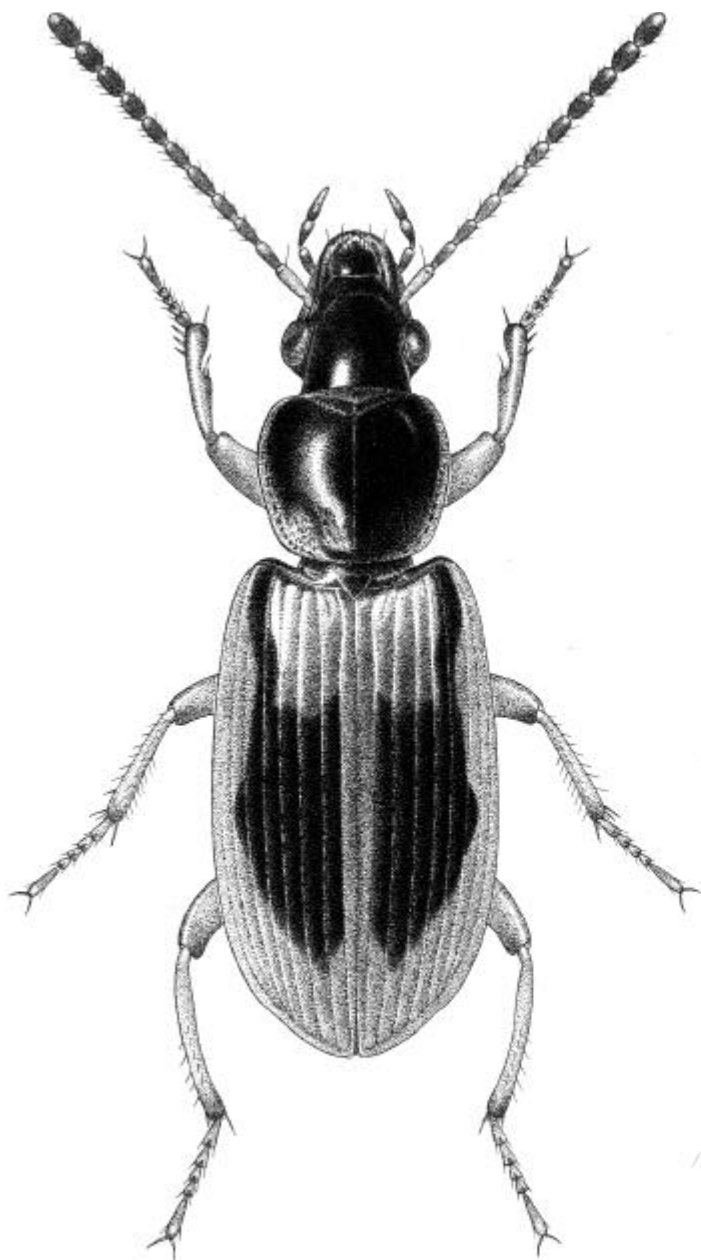


Fig. 1: *Acupalpus maculatus* Schaum (Carabidae). R.W.J. Read.

and *A. parvulus* are somewhat variable in the degree to which the pronotum is transverse. As noted by Hürka (1996), the pronotum of *A. maculatus* tends to be widest at the middle, whereas the pronotum of *A. parvulus* tends to have the widest point clearly anterior of the middle.

Depth and puncturation of basal foveae of pronotum

On average, British specimens of *A. maculatus* show slightly shallower pronotal foveae, and these with finer and sparser punctures than *A. parvulus*, but there is a degree of overlap between the two species.

Colouration of pronotum

The pronotum of *A. maculatus* averages more extensively dark than *A. parvulus*, with narrower pale borders. However, variation in both species means that there is considerable overlap in this character.

Male genitalia

A. maculatus has a smaller penis than *A. parvulus* (Fig. 2) and in dorsal view, the penis of *A. maculatus* is slightly narrower and in lateral view, less curved, than *A. parvulus*.

The male genitalia of both species are also illustrated by Jeannel (1941). The most distinct difference is apparent on clearing to examine the internal structures; *A. maculatus* lacks any chitinised internal structure, whereas *A. parvulus* shows 7 – 9 large chitinised teeth. It should be noted that according to Freude *et al.* (1976), *A. maculatus* possesses a single large chitinised internal tooth, in contradiction to Jeannel (1941) and to my observations of Dungeness specimens.

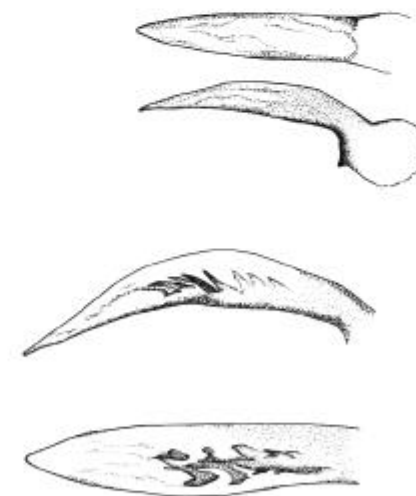


Fig. 2: Penis of *Acupalpus maculatus* top in dorsal and lateral views, and of *A. parvulus* below showing the numerous internal spines characteristic of the latter species.

Male pro-tarsi

The sexes of *A. maculatus* are comparatively difficult to separate, males having the pro-tarsi only feebly dilated compared with females, whereas in *A. parvulus* the males have more distinctly dilated pro-tarsi.

Prosternum

A. maculatus has a glabrous prosternum according to Jeannel (1941), differing from *A. parvulus* which Jeannel (1941) keys out as having a pubescent prosternum. However, Hürka (1996) appears to be correct in keying out both species as having 'prothorax and anterior abdominal sternites glabrous'.

HABITAT

According to Reitter (1908), *A. maculatus* inhabits salt-producing soils ("Auf salzhaltigem Boden", (salt-pans?)) in Germany. Freude *et al.* (1976) report only that it is a halophile. Hürka (1996) reports that in the Czech and Slovak republics *A. maculatus* is found "in moist, not shaded habitats: saline habitats, edges of waters with vegetation, lowlands".

At Dungeness, *A. maculatus* has been found at the edges of pools in quite dense vegetation, composed of rushes, mosses and other marsh vegetation. The Dungeness gravel pits are renowned for beetles of newly-created gravel pits with actively eroding margins devoid of vegetation, this being the favoured habitat of *Omophron limbatum* (Fabricius), *Dyschirius obscurus* (Gyllenhal), *Bembidion coeruleum* Serville, *Bembidion pallidipenne* (Illiger) (Carabidae) and *Heterocerus hispidulus* Kiesenwetter, (Heteroceridae) to name some of the rarest species. The discovery of *A. maculatus* suggests that later-successional vegetated gravel pit margins are equally worthy of investigation and of conservation management.

BRITISH RECORDS AND STATUS

All the confirmed British records that I am aware of are from Dungeness, VC 15 (Table 1). There have been no confirmed records of *A. parvulus* from the Dungeness area, and in retrospect, a record of *A. parvulus* from Dungeness ARC pits, TR073192 on 12 August 1999 by Peter Hodge is highly likely to refer to *A. maculatus* though no specimen was kept.

In 2002, after the initial announcement of the discovery of *A. maculatus* on the 'beetles-britishisles' e-mail discussion group, there were further records from the same area of Dungeness on 24 April (Tony J. Allen) and 3 September (Norman F. Heal), and quite probably there have been other records of which I am not aware.

It seems likely that *A. maculatus* has recently colonised Britain from the near continent. No further British specimens of *A. maculatus* have been located in the collections of either the Oxford University Museum of Natural History or the Natural History Museum London. Colonisation has probably occurred through natural dispersal across the English Channel, as also presumed for other recent arrivals at Dungeness: *Nebrioporus canaliculatus* (Lacordaire) (Dytiscidae) (Carr, 2000) and *Bembidion coeruleum* Serville (Carabidae) (Telfer, 2001). Though it appears to have

been established at Dungeness for at least seven years, it is still only known from quite a small area.

Table 1: Details of all known British records of *Acupalpus maculatus* up to the end of 2001, in chronological order.

Date	Grid ref.	Locality	Collector	Determiner	Number
15.ix.1996	TR069194	Dungeness, opposite side of road from the RSPB reserve	Norman F. Heal	Norman F. Heal, confirmed by MGT	1
2.x.1996	TR069194	Dungeness, opposite side of road from the RSPB reserve	Norman F. Heal	Norman F. Heal, confirmed by MGT	3
13.vi.1999	TR066202	Dungeness, near Boulderwall Farm	MGT	MGT	1
8.v.2000	TR069189	Dungeness	Peter Whitton	Peter Whitton	3
18.v.2000	TR0620	Dungeness	MGT	MGT	3
18.v.2000	TR0719	Dungeness	Eric G. Philp	Eric G. Philp, confirmed by MGT	6
18.v.2000	TR0620	Dungeness	Eric G. Philp	MGT	2
21.vi.2001	TR066199	Dungeness	John Paul	MGT	3
12.x.2001	TR06561989	Dungeness	Peter J. Hodge	Peter J. Hodge	1

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I thank James Hogan of the Oxford University Museum of Natural History for checking their British specimens of *A. parvulus*. I also thank Martin Brendell and Max Barclay at the Natural History Museum for arranging access to the collections there. I am indebted to John Paul, Peter Whitton, Peter Hodge, Eric Philp, Norman Heal and Tony J. Allen for details of their records of *A. maculatus*, without which this paper would be incomplete. John Paul and Peter Whitton both helped with the preparation of the manuscript. Simon Busuttill of the RSPB deserves thanks for welcoming coleopterists to the Dungeness RSPB reserve.

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More recent *Philhygra terminalis* (Gravenhorst) (Col. Staphylinidae) records

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The note of this species occurring in Surrey and Leicestershire during 2002 (Lott, 2002), although quoting recent records from Hyman (1994), fails to acknowledge recently published records (as *Atheta terminalis* (Gravenhorst)) from Yorkshire (Denton, 1996 & 2000). These specimens, from Wheldrake Ings (VC 61) on 11 March 1995 and 25 October 1996, were collected from flood refuse and *Glyceria* litter respectively. Despite extensive searching by the likes of Colin Johnson, Bob Marsh and the author, these were the first county records since 1933.

The fact that Lott (*op. cit.*) recorded *P. terminalis* with its congener, *P. gyllenhali* (Thomson), is of interest, as on both the above-mentioned dates the two species were also found together.

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Pelenomus olssoni Israelson (Curculionidae) in Kent

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I attended a meeting of the Kent Field Club at the R.S.P.B. Tudeley Woods Reserve, Pembury VC 16 on 29 July 2001. It was extremely hot (31° C), very few beetles were about and after 5½ hours trudging around some 4 km of the reserve, exhausted I rested by the edge of an almost dried up pond at TQ6141 Newbars Wood.

Noticing a welcome movement amongst the prostrate vegetation, I extracted a few beetles including a *Pelenomus* sp. that upon later dissection proved to be a male *P. olssoni* Israelson. Prof. M.G. Morris very kindly confirmed my identification and commented that it appears to be the first record for the county of Kent. A subsequent revisit under more favourable weather conditions enabled me to use a mechanical suction device, finding the species to be plentiful.

[The Coleopterist **12**(1): 1-6 April 2003]

Ceutorhynchus cakilis (Hansen) in the British Isles (Curculionidae)

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Introduction

Although records of this species have been published (Owen, 1990; 1993; Morris, 1992; 1993) no detailed and comprehensive account of its occurrence in the British Isles has been given. In this short paper the records are amplified, and comments on the nomenclature, identification, biology and distinctness of the species are made.

Systematics and Identification

Ceutorhynchus cakilis was described by Hansen (1917) (under *Ceuthorrhynchidius* (*sic*)) as a good species 'though very closely allied to' the common *C. floralis* (Paykull, 1792) (nec (Olivier, 1790); now *C. typhae* (Herbst, 1795) (Colonnelli, 1993). This status was reduced to that of a form in Hansen (1965). It was included as such in Hoffmann (1954), as a 'race'. However, the taxon was accepted as a good species by Dieckmann (1972) on the basis of new information ('neuer Erkenntnisse') which modified his earlier opinion (1966) that it was only a variety of *C. floralis*. The taxon's status as a good species has been followed by subsequent authors (e.g. Smreczynski, 1974; Lohse, 1983; Tempère & Péricart, 1989; Silfverberg, 1992).

C. cakilis, *C. typhae* and their allies (species of *Ceutorhynchus s.str.* with a six-segmented antennal funiculus) are often placed in a subgenus *Neosirocalus* Wagner, 1944 (Dieckmann, 1972; Lohse, 1983). Tempère & Péricart (1989) and Dieckmann (1966) attributed the name to Neresheimer & Wagner, 1938, but this name is invalid as no type-species was designated. However, subgenus *Neosirocalus* Wagner (which is in any case a synonym of *Ceuthorrhynchidius* Jacquelin du Val, 1855) is not accepted by Colonnelli, the world authority on Ceutorhynchinae (in Alonso-Zarazaga & Lyal, 1999). Species of *Ceutorhynchus* with a six-segmented antennal funiculus are probably not a monophyletic group, and subgeneric names for the group have had little currency in British literature since the time of Fowler (1891). However, the biology of species in the group is similar, larvae feeding in the fruits of brassicaceous plants and pupating in soil; the shape of the male median lobe is also similar (Dieckmann, 1966).

C. cakilis may be distinguished from other species of *Ceutorhynchus* by the following key:

- | | |
|---|--------------|
| 1 Antennal funiculus with seven segments. | [other spp.] |
| - Antennal funiculus with six segments. | 2 |

[The Coleopterist **12**(1): 7-13 April 2003]

2 Pronotum with a distinct, sharp tubercle on each side, about one-third from base. 3

- Pronotum without sharp tubercles (though with raised, blunt prominences on each side in some cases). [other spp.]

3 Elytral interstices mostly with two rows of scales or broad setae; on average smaller and less robust, 1.5 - 2.4 mm [very common species; on a wide range of Brassicaceae]. *typhae* (= *floralis*)

- Elytral interstices mostly with three rows of scales or broad setae; on average larger and more robust, 2.2 - 2.6 mm [rare coastal species; on *Cakile maritima* Scop. and possibly *Crambe maritima* L. only]. *cakilis*

The only other character given by Hansen (1917) and subsequent authors to distinguish *cakilis* from *floralis* (*typhae*) is the broader and more conspicuous sutural scales of the former taxon. However, this character would appear to be variable within the *typhae-cakilis* complex.

Keys to species of '*Neosirocalus*' were provided by Dieckmann (1972) and by Lohse (1983), but neither includes *C. hepaticus* Gyllenhal, a rare British species. Dieckmann's more comprehensive key to the species of Central Europe (1966) includes both *C. hepaticus* and *C. cakilis* (as a form of *floralis*). The male median lobes of *C. cakilis* and *C. typhae* do not seem to differ significantly, but assessment is handicapped by lack of material.

***C. cakilis* in the British Isles**

The first record of the species in the British Isles is that of Owen (1990). He exhibited a pair of the weevil, which he had taken on a coastal site near Grenitote, North Uist, Outer Hebrides, in July 1979, at the meeting of the British Entomological and Natural History Society on 25 April 1990. He provided notes on the specific identity, recognition and host plants. He gave the size of *C. floralis* as 1.5-2.1 mm and this is indeed the normal range, but, exceptionally, larger examples up to 2.4 mm are occasionally found, as noted in the key above. Owen's record was repeated, without details, by Owen (1993). In this publication the page reference to the original record is given incorrectly as 81; it should be 181. The site is in vice county 110 at NGR NF8277, and is named Machair Leathann on the 1:50,000 O.S. map (sheet 18).

C. typhae (as *floralis*) has been recorded from both the Outer and Inner Hebrides. Records from North and South Uist are included by Waterston *et al.* (1981); the record from North Uist could refer to JAO's specimens of *C. cakilis*. Welch (1983) recorded *C. floralis* from Canna, and Wormell (1982) included a record of that species taken by MGM on Rum (this specimen has been re-checked and is clearly *typhae*).

Morris (1992) recorded a single female of *C. cakilis* at Five Mile Point, Co. Wicklow, Ireland, on 18 July 1991; this record was repeated by Morris (1993). A further female was taken at the same site (vice-county H20, IGR 03102) on 31 May 1995. Both specimens were on *Cakile maritima*. The site is a lengthy, but narrow, strip of coastal sand and notable for supporting other rare Irish weevils, for example *Diplapion confluens* (Kirby) and *Tychius squamulatus* Gyllenhal (Morris, 1997).

The four examples mentioned have been compared with each other and are the only ones so far recorded from the British Isles, but it is not improbable that large specimens of *C. typhae* taken on the coast will be found to be actually this species, particularly in view of its European distribution and abundance. JAO has given his two specimens for incorporation into MGM's collection, so that the four weevils may be kept together.

Biology

Dieckmann (1972) gave a fairly detailed account of the life history of *C. cakilis* in Germany. Because *Cakile maritima* does not bloom there before July, oviposition did not occur at the earliest until the end of that month. Larvae were found in the fleshy pods of the host in early September; they fed only on the seeds and did not attack the pericarp. Larvae pupated in mid September, and adult beetles started to emerge at the beginning of October.

Cakile maritima is a plant of somewhat uncertain or erratic occurrence. It can be abundant at times and at others scarce or absent, even at the same site. This may affect the presence and abundance of the weevil. *Cakile maritima* is pre-eminently a coastal, or maritime, species (Fig. 1).

Crambe maritima has been recorded as a rarer host of *C. cakilis* but there are no details of its biology on that plant, if indeed it is a larval foodplant. An up-to-date map of its distribution in the British Isles is given in Preston, Pearman & Dines (2002).

European distribution of *C. cakilis*

The weevil has been recorded from Fennoscandia: Sweden, Norway and Denmark (Silfverberg, 1992), from East and West Germany and Poland (Lucht, 1987), from northern France (Pas-de-Calais) (Hoffmann, 1954) and from Sicily (Abbazzi & Osella, 1992). The last record is remarkably far from others. *C. cakilis* has not been recorded elsewhere in the Mediterranean region and it is possible that some specimens from Sicily that were examined by Dieckmann (1966) were not *C. cakilis* (*vide infra*); however, Dieckmann (1972) included the island in his summary of the species' distribution. In all countries, *C. cakilis* is restricted to coasts, because of its dependence on its strictly maritime hosts. Thus, in Poland it has been recorded only from the one area of the Baltic Coast (Burakowski, Mroczkowski & Stefanska, 1997), whereas in Denmark it is widely distributed, having been recorded in all divisions of the country except North West Jutland (Hansen, 1996).

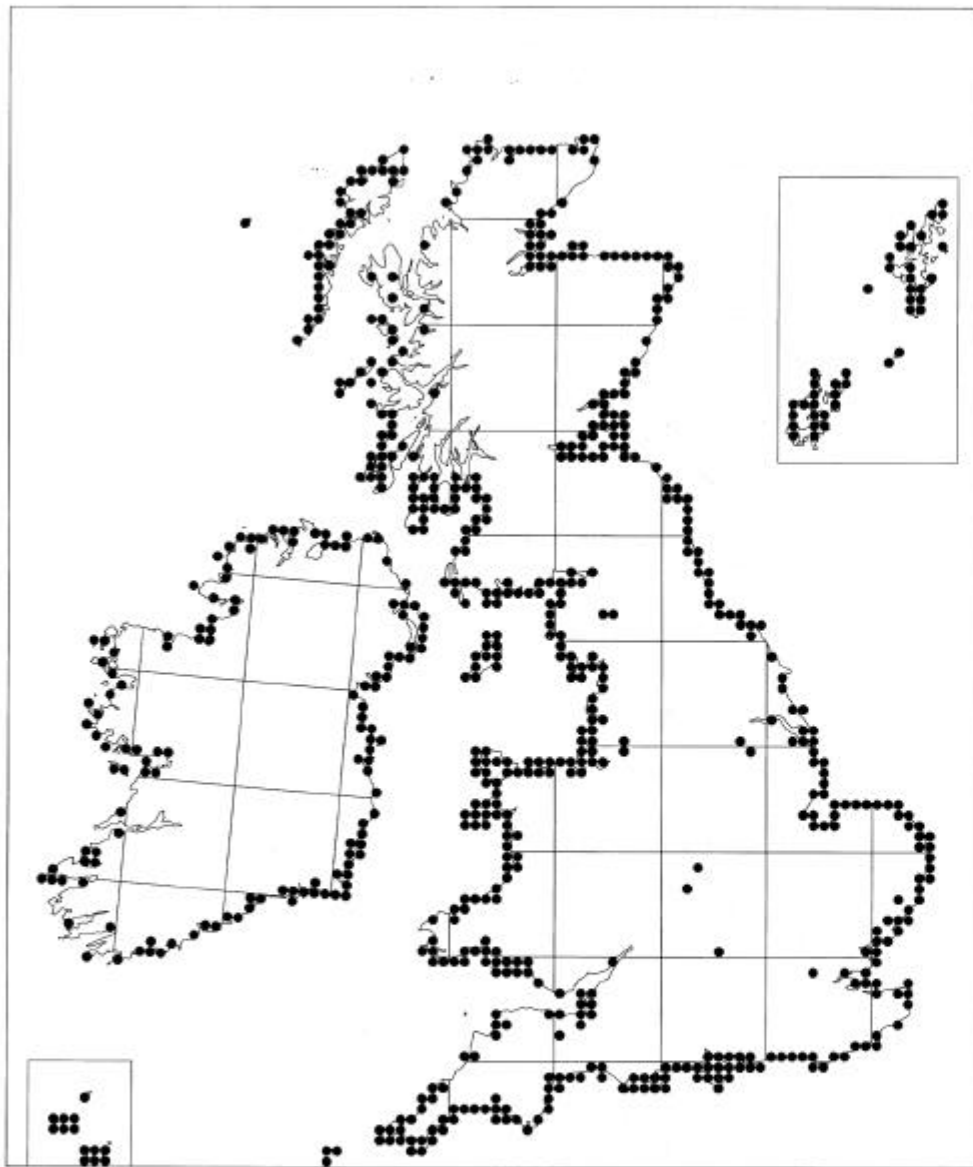


Figure 1. 10 Km square distribution of *Cakile maritima* in the British Isles. Reproduced by permission of the Biological Records Centre, Centre for Ecology and Hydrology, Monks Wood.

Specific distinctness of *C. cakilis*

Most recent authors have followed the expertise of Dieckmann (1972) in recognising this taxon as a good species. However, it is very close to *C. typhae*, and mention must be made of the possibility that the larger size is correlated with the additional row of setae on the elytral interstices. A possibly analogous (though reverse) case is that of British '*Miarus degorsi* Abeille', (Owen, 1988; 1993). It was concluded that examples of this taxon, being smaller than *M. graminis* Gyllenhal (with which it has been tentatively regarded as conspecific), could have the number of rows of interstitial setae reduced. Further details may be found in Owen's papers. It is worth pointing out that the late Dr Lothar Dieckmann suggested, or at least agreed, with this possibility (*in litt.*).

It is clear that the specific distinctness of *C. cakilis* has been doubted. Hansen (1965) evidently revised his earlier assessment of the taxon as a good species (Hansen, 1917). Both Horion (1935) and Wagner (1944) suggested that it might be a foodplant race of *C. floralis*, though both included it as a good species. Dieckmann's (1966) assessment of the taxon as a variety of *C. floralis* may have been influenced by the opinion of Smreczynski (1955), but perhaps more by his receipt of specimens collected in Sicily. Some of these were taken from *Rorippa nasturtium-aquaticum* (L.) and would appear to be merely large examples of *C. typhae*, rather than true specimens of *C. cakilis*, which is associated only with *Cakile* and *Crambe*. This is probably the basis for Dieckmann's revised opinion (1972) of the taxon's status, an opinion evidently shared by Smreczynski (1974), revising his earlier assessment (1955). All authors quoted emphasised the association of *C. cakilis* with *Cakile maritima* (and possibly *Crambe maritima*).

Several forms of *C. typhae* have been described, although the only British one that has been raised to specific level is *C. palustris* (= *Ceuthorrhynchidius palustre*) (Edmonds, 1930; Donisthorpe, 1931). It is currently regarded as a small form of *C. typhae* (at best), is possibly a foodplant race associated with *Rorippa nasturtium-aquaticum*, and is also recognised in France. Hoffmann (1954) attributed the description erroneously to Edwards, and Tempère & Péricart (1989) ascribed the name of the 'morphé' to Hoffmann himself.

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Records of four carabids potentially new to Dorset

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From August 1977 to February 1978, TH sampled for carabid and staphylinid beetles on several heathlands in Dorset, while based at the then Institute of Terrestrial Ecology's Furzebrook research station. Most sampling was done by pitfall trapping (with over 97,000 trap days in total), supplemented by hand-searching. The total number of specimens caught and identified was 32,571 including 20,020 adult carabids (Heijerman, 1978). Among the 119 carabid species recorded are four that are potential additions to the Dorset list. These four are not listed for Dorset by Moore (1957) in his tabulation of the county distributions of British carabids, neither A.J. Allen nor Prof. M.G. Morris (pers. comms.) are aware of any Dorset records, and there are no Dorset records in Luff (1998) or subsequently reported to the Ground Beetle Recording Scheme.

Cicindela hybrida Linnaeus

Hartland, three females in pitfall traps in heathland collected on 16 September 1977, one at ST9485 and two at SY9585.

Bembidion octomaculatum (Goeze)

Cranborne (c. SU1011), 9 August 1977, one female collected by hand along a ditch.

Bradycellus collaris (Paykull)

Parley (SZ0999), two males in pitfall traps collected on 8 December 1977 from a heathland site. Holt (SU0603), one male in pitfall traps collected on 8 December 1977 from a heathland

site. Middlebere (c. SY9484), one female in pitfall traps collected on 3 November 1977 from a dry woodland site on the edge of a meadow. Ridge (SY98), one female in pitfall traps collected on 6 November 1977, habitat unrecorded.

Amara famelica Zimmermann

Holt (SU0603), one male in pitfall traps collected on 8 September 1977 from a heathland site. Arne (c. SY9687), one female in pitfall traps collected on 15 September 1977 from a heathland site.

Records of these species from Dorset are plausible based on current knowledge of their British distributions and habitat. It would be desirable to check specimens to confirm the records, but unfortunately, no specimens are available – they were kept until 2000, but thrown out in that year (R. Snazell, pers. comm.). Therefore, these are unconfirmed additions to the Dorset list.

The most surprising of these potential additions is *C. hybrida*, which seems unlikely to have been overlooked in Dorset. *Bradycellus collaris* is a much less conspicuous species, and one that could be overlooked or misidentified. *Bembidion octomaculatum* and *A. famelica* are both very rare species in Britain, with a tendency to be recorded rather sporadically, as single specimens, (though *B. octomaculatum* has also been recorded in good numbers in Britain (Jones, 1992)). All these records should provide a spur to coleopterists to make further searches.

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Rare and notable Coleoptera from coastal sites in north-east England

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Introduction

Since 1985 we have sampled a number of sites for beetles along the north-east England coast, covering dunes, slacks and saltmarshes. The records of several of the nationally scarce species are in Eyre & Luff (1987) and Luff, Eyre & Jessop (1996) but most records have not been reported. The published records are also included here for completeness. The distribution of carabid, staphylinid and phytophagous beetle assemblages on the north-east England coast are discussed in Eyre & Luff (2002).

Sites

The southernmost site sampled was Coatham Sands (NZ5725), just south of the mouth of the River Tees, VC 62, North-East Yorkshire, in 1995. Coatham Sands has a mixture of habitats with loose and fixed dunes, slack and three ponds. In Co. Durham, VC 66, the North Gare Sands (NZ5327), loose dunes and slack were sampled in 1996, as were the loose and fixed dunes of Seaton Sands (NZ5328), and saltmarsh sites by Greatham Creek and on Cowpen Marsh (both NZ5025) were sampled in 1992 and 1996 respectively. In Northumberland VC 67, saltmarsh at Seaton Sluice (NZ3376) was sampled in 1996, whilst loose and fixed dune and slack areas at Druridge Bay (NZ2894) were sampled in 1995 and 1996. In VC 68, saltmarsh and fixed dune at Alnmouth (NU2409), saltmarsh at Beal (NU0842) and Budle Bay (NU1434) and saltmarsh (NU1043) and dune and slack sites (NU0943) on Holy Island were all sampled in 1996.

Sampling was carried out by pitfall traps at all sites, whilst the aquatic sites at Coatham Sands were sampled with a pond net. National rarity and statuses follow those given in Hyman, 1992, 1994.

Rare and notable species

APIONIDAE

Oxystoma cerdo (Gerstaecker) **Nb**

Recorded from fixed dune at Druridge Bay in 1996. Dunes are not mentioned as a habitat by Hyman (1992). There are recent records from spoil heaps and disturbed grassland in north-east England (Luff, Eyre & Jessop, 1996).

CARABIDAE

Amara fulva (Müller) **Nb**

Found on a number of sandy riverine sediments (Eyre, Luff & Lott, 2000), this species was recorded on both loose and fixed dunes at Druridge Bay in 1995 and 1996.

Amara lucida (Duftschmid) **Nb**

This mainly coastal species was found on the loose and fixed dunes at Coatham Sands in 1995, and on the loose and fixed dunes at North Gare and Seaton Sands in 1996. These records are the most northerly on the eastern coast of Britain (Luff, 1998).

Amara praetermissa (Sahlberg) **Nb**

There is a record for this species from Seaton Sands (NZ5229) by M. Denton in the 1980s (Luff, Eyre & Jessop, 1996) and more from colliery spoil in the region (Eyre & Luff, 1995).

Amara spreta Dejean **Nb**

Found on both loose and fixed dunes at Coatham Sands in 1995 and Seaton Sands in 1996, these are the only recent records on the east coast of Britain north of Kent and are the most northerly British records (Luff, 1998).

Bembidion clarki Dawson **Nb**

Found in the wet slack areas of Coatham Sands in 1995 and North Gare Sands in 1996, there is only one more northerly record in Britain (Luff, 1998).

Bembidion laterale (Samouelle) **Nb**

A saltmarsh species, it was recorded from this habitat at Beal and Holy Island in 1996.

Bembidion pallidipenne (Illiger) **Nb**

This species was recorded from wet streamside sand in the dune system at Druridge Bay in 1985 (Eyre & Luff, 1987). The only other recent records on the British east coast are Scottish (Luff, 1998).

Calathus ambiguus (Paykull) **Nb**

Recorded from the loose dunes at Coatham Sands in 1995 and the fixed dune at Seaton Sands in 1996, this is the most northerly recent British record (Luff, 1998).

Dyschirius impunctipennis Dawson **Nb**

This species frequents sandy places by the coast (Hyman, 1992) a habitat found on the saltmarsh at Budle Bay, from where it was recorded in 1996. This is again the most northerly recent record in Britain (Luff, 1998).

CHRYSOMELIDAE

Mantura rustica (Linnaeus) **Nb**

A species of sandy soil sites found on sandy riverine sediment by the River Tyne (Eyre, Luff & Lott, 2000) and recorded from a fixed dune at Druridge Bay in 1996.

COCCINELLIDAE

Scymnus schmidti Fuersch **Nb**

Also found on riverine sediment, by the River Nith in southwest Scotland (Eyre, Luff & Lott, 2000), it was found on fixed dune on Holy Island in 1996. North-east England is not given in the distribution information of Hyman (1992) and it may be new to the region.

CURCULIONIDAE

Barynotus squamosus Germar **Nb**

A species of well-drained soils, there are several records for this species from inland sites in Eyre, Luff & Lott (1998, 2000) and it was found on the coast by Greatham Creek in 1992.

Cleonus piger (Scopoli) **Nb**

This species feeds in the stems of creeping thistle *Cirsium arvense* (L.), which grows on the fixed dune sites at Druridge Bay and Holy Island, from where it was recorded in 1995 and 1996.

Glocianus punctiger (Sahlberg) **Nb**

Apparently mainly associated with coastal sand sites (Hyman, 1992), it was found on fixed dune at Druridge Bay in 1996.

Grypus equiseti (Fabricius) **Nb**

There are many pitfall trap records for this species on inland sites (Eyre, Luff & Lott, 1998, 2000) and it was found on fixed dune and slack areas of Coatham Sands in 1995, on loose dune and slack areas of North Gare Sands in 1996 and on slack areas of Druridge Bay, also in 1996.

Omiomima mollina (Boheman) **Na**

A species previously found on well-drained grassland and inland riverine sediment sites (Luff, Eyre & Jessop, 1996; Eyre, Luff & Lott, 2000), it was recorded from the Holy Island saltmarsh area in 1996.

Otiorhynchus desertus Rosenhauer **Nb**

There is an old record for this species from Newton Links on the Northumberland coast and from a spoil heap (Luff, Eyre & Jessop, 1996), and it was found on a number of riverine sediments (Eyre, Luff & Lott, 2000). It was recorded from fixed dune sites at Alnmouth and on Holy Island in 1996.

Otiorhynchus raucus (Fabricius) **Nb**

Not, apparently, a coastal species (Hyman, 1992), it was found by Greatham Creek in 1992 (Luff, Eyre & Jessop, 1996).

Pelenomus zumpti (Wagner) **Na**

Previously found in southwest Scotland by the upstream River Nith (Eyre, Luff & Lott, 1998), a record from Seaton Sluice saltmarsh in 1996 is likely to be the first for northeast England (Hyman, 1994).

Phyllobius vespertinus (Fabricius) **Nb**

This species is apparently found mainly on saltmarshes (Hyman, 1992; Ball, 1997) but it was found on fixed dune at Coatham Sands in 1995 and on loose dune at North Gare Sands and fixed dune at Seaton Sands, Druridge Bay and Alnmouth, all in 1996. There were no records from saltmarsh sites.

DYTISCIDAE

Ilybius subaeneus Erichson **Nb**

This species was found in the ponds on Coatham Sands in 1995.

GEOTRUPIDAE

Geotrupes mutator (Marsham) **Nb**

Found on the slack area at Druridge Bay in 1996, there was plenty of horse dung on the adjacent pasture in which it could have developed.

HALIPLIDAE

Halipus apicalis Thomson **Nb**

Recorded in the Coatham Sands ponds in 1995, this species was also found in the pitfalls situated in the slack area of North Gare Sands, near a small pond, in 1996.

HELOPHORIDAE

Helophorus fulgidicollis Motschulsky **Nb**

Usually found on saltmarshes, this species was also recorded from the pitfalls on the slack area at North Gare Sands in 1996.

HYDRAENIDAE

Ochthebius bicolon Germar **Nb**

There were a number of records of this species from riverine sediments in Eyre, Luff & Lott (2000) and it was recorded from saltmarsh sites at Cowpen Marsh, Seaton Sluice, Alnmouth, Beal and Holy Island and from the dune slack at North Gare Sands, all in 1996.

HYDROPHILIDAE

Cercyon tristis (Illiger) **Nb**

A wetland species, it was recorded from the dune slack areas of Coatham Sands in 1995 and North Gare Sands in 1996, as well as from Cowpen Marsh in 1996.

Cercyon ustulatus (Preyssler) **Nb**

Another wetland species, it was found in the Druridge Bay slack area in 1996.

Enochrus bicolor (Fabricius) **Nb**

This saltmarsh species was recorded from Cowpen Marsh in 1996.

LEIODIDAE

Hydnobius punctatus (Sturm) **Notable**

Recorded from a chalk spoil heap in Northumberland and on the loose dunes at Coatham Sands in 1995 (Luff, Eyre & Jessop, 1996).

Leiodes ciliaris (Schmidt) **Notable**

Recorded from loose dunes at Coatham Sands in 1995 (Luff, Eyre & Jessop, 1996), it was found on North Gare Sands in 1996, also on loose dunes.

Trichohydriobius suturalis (Zetterstedt) **RDBK**

Found on loose dunes at Coatham Sands in 1995 (Luff, Eyre & Jessop, 1996).

SCARABAEIDAE

Euheptaulacus villosus Gyllenhal **Na**

Found on loose dunes at North Gare Sands, this record comes between recent records from southwest Yorkshire and Angus (Hyman, 1992).

SILPHIDAE

Aclypea opaca (Linnaeus) **Na**

Recorded from woodland, a cultivated field, an offshore island and a number of riverine sediments (Luff, Eyre & Jessop, 1996; Eyre, Luff & Lott, 2000), it was found on fixed dune at Druridge Bay in 1996.

STAPHYLINIDAE

Aleochara ruficornis Gravenhorst **Notable**

Another species found in a number of habitat types (Eyre, Luff & Lott, 1998, 2000), it was found on the coast by Greatham Creek in 1992.

Diglossa submarina (Fairmaire & Laboulbène) **Notable**

Found on the loose dunes near the high tide limit on Coatham Sands in 1995.

Gabrius bishopi Sharp **Nb**

A species recorded from a range of habitat types (Luff, Eyre & Jessop, 1996; Eyre, Luff & Lott, 2000), it was found on fixed dune at Druridge Bay in 1995 and on the slack area at North Gare Sands in 1996.

Gabrius keysianus Sharp **Nb**

Previously found on coastal sites, this species was recorded from a slack site on Holy Island in 1996. North-east England was not given in the distribution information in Hyman (1994) and it may be a new record for the region.

Gabrius osseticus (Kolnati) **Nb**

A wetland species usually found near the coast (Hyman, 1994), it was found on the slack area of Coatham Sands in 1995.

Mycetoporus piceolus Rey **Notable**

Recorded from dry chalky or sandy sites, this species was found on fixed dune at Coatham Sands in 1995.

Ocypus nero (Faldermann) **Na**

There are recent records of this species from riverine sediments (Eyre, Luff & Lott, 2000), it was also found on the saltmarsh at Holy Island in 1996.

TENEBRIONIDAE

Crypticus quisquilius (Linnaeus) **Nb**

A coastal species, it was recorded from fixed dunes at Alnmouth and Druridge Bay in 1996. The distribution for this species is given as north to the Humber/Mersey by Ball (1997), in contrast to the information given by Hyman (1992) that it has been recorded from northeast England and southwest Scotland.

Scaphidema metallicum (Fabricius) **Nb**

Recorded from a variety of habitats, including those on the coast (Hyman, 1992), it was found on fixed dunes at Seaton Sands in 1996.

Discussion

There was a marked difference between the lists of rare and scarce beetle species recorded from the dunes at Teesmouth and those of the Northumberland coast. Similar numbers of rare and scarce species were recorded from Coatham (13), Seaton/North Gare Sands (15) and at Druridge Bay (13) but only one coastal species, *Phyllobius vespertinus*, was common to the dune sites of the two areas. Only two other non-specific habitat species, *Gabrius bishopi* and *Grypus equiseti*, were found on both Teesmouth and Northumberland dunes. This pattern is also seen in the records from the saltmarshes. The only species common to the two areas was *Ochthebius bicolon*, not a species limited to saltmarshes.

The dune systems at Teesmouth differ in size, structure and, probably, composition from those in Northumberland of Druridge Bay, Alnmouth and Holy Island. The loose dunes at Coatham and North Gare Sands have greater depth than those in Northumberland but the greatest differences are in the fixed dunes. The fixed dunes at Coatham and Seaton Sands are several hundred metres wide whilst those on Druridge Bay are less than 50m. The Alnmouth and Holy Island fixed dunes are more extensive but they have far more dense, scrubby vegetation than those at Teesmouth, where they are open with ruderal vegetation. The sand appears to be finer at Teesmouth than in Northumberland, probably because it is derived from the river whilst the coarser sand on Druridge Bay, Alnmouth and Holy Island is likely to be a product of sea action with a higher shell content.

Within habitat classifications of ground, rove and phytophagous beetles (Eyre & Luff, 2002), the sandy dune, slack and saltmarsh sites of Teesmouth and Northumberland tended to be in separate groups. The majority of common species occurred on the dunes (e.g. *Calathus fuscipes* (Goeze), *C. mollis* (Marsham)) and saltmarshes (e.g. *Dicheirotichus gustavi* Crotch) all along the coast. The differences were amongst the rarer species and, whilst a number of species will be common, it may be that the quality of other dune and saltmarsh sites for beetles depends on their structure and origin.

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Some Entomological Recollections of Baron Alexis de Porochin (1918-1980)

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One of the outstanding members of the Manchester Entomological Society when I joined about 1950 was Baron Alexis de Porochin, a small but flamboyant individual with whom I struck up an immediate friendship, aided by his extremely kind and helpful manner. He was to form a pivotal role in my career, as it was through him that I switched from a planned career in art to one in entomology. A brief biographical account of Alec, along with a portrait of him as he was when we first met, appears in Underwood (1996)

In the above-mentioned published biography it is noted that Alec was of above average ability as a field-worker, honed no doubt by his long association with the Finnish Entomological Society in Helsinki, where he clearly learnt the art well. His knowledge of the ecological requirements of many beetles was considerable, and, since many rarities in Britain are common in Finland, highly relevant to the British worker. Alec was for instance the only person I have known who had regularly taken *Lamia textor* (Linnaeus). Although the species was common in Finland, he recalled that its discovery in any particular area required considerable know-how and perseverance. He was of course aware of the crepuscular or nocturnal habits of the beetle, recalling that a good clue to its presence were small patches of freshly gnawed bark about the bases of *Salix* boles. These could be sought during the day and then revisited in the dark or half-light with a torch, when the beetles themselves could be seen, sometimes in large congregations.

Alec and I enjoyed very many collecting trips over the 11 years I worked with him and it was on one such trip, probably in June 1955, that we visited Mouldsworth Pond near Delamere in Cheshire. I was some distance away from him when a sudden torrent of Finnish, Swedish and German expletives emanating from the marshy swathe beside the pond indicated that Alec was displeased, so I went over to him. Calming down, he said he had just missed a *Bromius obscurus* (Linnaeus). We sought in vain for the specimen but no more were to be seen. Owing to the passage of time, I cannot be absolutely sure, but I have the vaguest recollection that he drew my attention to the characteristically slashed leaves indicating where the beetles had been feeding. In any case, I had no reason to doubt that Alec had seen the beetle on this occasion as he had spoken about the species many times, saying it was one of the commonest beetles in Finland. He recalled how it invariably fell to the ground when approached and hence often escaped notice or capture in the sweep-net. Anyway, the Mouldsworth event left me completely bewildered but slowly passed into the back of my mind.

Nearly thirty years later, at a meeting of the Entomological section of the Yorkshire Naturalists Union, Peter Kendall showed me a strange beetle from Danebridge near Congleton in Cheshire that had defied all his attempts at identification. I recognised it immediately as *Bromius obscurus* and his record was soon published (Kendall, 1982).

Although Alec's sighting of *Bromius obscurus* has remained unpublished until now, I have mentioned it several times over the years to coleopterists familiar with the Delamere area in the hope that the species may be re-discovered there and Alec's record thereby confirmed. However, I never had any feedback and found, on a lightning visit to the site several years ago, that the pond had been 'tidied up' and was much changed. In the 1950s when the sighting was made, the outlet stream from the pond was on its southern side, immediately south of a broad marshy expanse on the pond's western edge (SJ517710). There was a rich flora in this marsh, including a large patch of rosebay willowherb *Chamerion angustifolium* (L.) upon which Alec saw the beetle. In passing, it may be mentioned that Mouldsworth is about 35 miles west of Peter Kendall's site. Alec also recalled that, although the beetle is very common in Finland, it is not really ubiquitous but tends to occur in colonies, which however may be very populous. Professor Paul Buckland (pers. comm.) also found this to be the case in Swedish Lapland and northern Norway. I also gather from him that it was apparently one of the commonest beetles in Britain in early post-glacial times. Although the Mouldsworth site has been greatly altered since the mid 1950s, a search in the vicinity may prove successful.

The bulk of Alec's collection, including much Finnish material, passed eventually to Manchester Museum after his death in 1980. A smaller number were given to me beforehand and these were deposited in the Doncaster Museum. Recognition of Alec's specimens in collections could prove difficult, as he never included his name on data labels; as noted in his biography he was implacably opposed to such a form of self-advertisement as he saw it. His style of lettering however was characteristic, in extremely neat italicised hand, and his data labels consistently measured 5 by 13 mm. The name of the insect was usually written in the same style on the underside of the data label, typically followed by A de P. in the right lower corner. Alec above all else was a creature of habit.

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Gastrallus immarginatus (Müller) (Anobiidae) at Hatfield Forest, Essex

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For many years, this species was only known from Windsor Great Park and Forest and it still survives there in the very localised ancient field maple *Acer campestre* L. trees. It has however been found at a number of additional sites in recent years, notably along the Severn Vale, where it is scattered across a wide area of country in north Gloucestershire and south Worcestershire. Other recent reports that I am aware of are for Knole Park, Kent (Hance, 1998) and Hatchlands Park in Surrey (Denton & Alexander, 2002). It is also one of the chosen few of our many rare and threatened saproxylic beetles to grace the list of Priority Species in the UK Government's Biodiversity Action Plan (Anon, 1999).

I can now report its discovery in a sixth county, Essex. One of the many special features of Hatfield Forest in N. Essex (VC19) is its many ancient field maple pollards and so, of all UK sites, this really is where this beetle should be found. A special concerted effort made in 2002 was very successful. The characteristic exit holes and galleries were found to be present on ancient open-grown field maple pollards scattered through the Central Plain. It was found on a total of seven trees in the 1km squares TL5318, 5319 and 5320, on 31 July, and 1 and 6 August.

Interestingly none could be found on ancient field maple pollards on the more intensively recorded Takeley Hill and Bushend Plains (TL5420) of the Forest. It may be relevant that these latter areas were actually ancient coppices which were abandoned by the early 17th century (Rackham, 1989), opened up to grazing, and surviving trees converted to pollards. Clearly the species has been so immobile in the Forest that it has not been able to cross from the Central Plain to colonise these unoccupied areas barely ½ km away to the east, and within a period of about 400 years. It is perhaps significant that other old coppices – now largely closed canopy woodlands with only relatively young aerial growth of field maple – occupy most of the intervening land, and presumably form an effective barrier to movement of this species.

Acknowledgements

Many thanks to Vikki Forbes, former Property Manager at Hatfield Forest, for stimulating the visit.

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Otiorhynchus setosulus Stierlin 1861- (Curculionidae) as a British species

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On 14 August 1992 – a cold windy day, I had beaten several *Otiorhynchus* (*Liophloeus*) specimens from a short length of old ivy-covered hawthorn hedge at Darenth, north-west Kent (TQ5771). Catching up on identification a couple of years later, I realised that one of the *Otiorhynchus* weevils looked very different. It was duly despatched to Prof. M.G. Morris who readily offered to look at the specimen. After examining specimens in the Natural History Museum General Collection, (there were four examples – three with data from Italy), he was able to determine it as *O. setosulus* Stierlin in time to include reference to the species in the 1997 Royal Entomological Society handbook, Broad Nosed Weevils (Morris, 1997). He recorded it as 'a single specimen.....probably introduced'.

Stierlin's (1861) account of European *Otiorhynchus* species records *O. setosulus* from Sicily. The distribution is extended by Lona (1936) to include also central/southern Italy, and the imagination was duly exercised as to why with such restricted distribution, this flightless weevil should suddenly appear in north-west Kent.

At about the same period, I was cataloguing the Coleoptera collection of the late K.C. Side in the Maidstone Museum of Art and Natural History, and I was very surprised to find a second *O. setosulus* in a cabinet drawer of principally foreign holiday species labelled 'Stone, Kent – TQ57 - 7.6.1958'. Fortunately, his field diaries still survive and confirm the data are correct, the specimen having been identified by Mr. R.T. Thompson (Natural History Museum, London) in 1959 and thought to be a casual importation from the Mediterranean region. Stone is located a couple of miles to the north of Darenth.

Finally Richard Jones informed me that he had identified a specimen as *O. setosulus* beaten from wild cherry *Prunus avium* (L.) at Elm Park nr Havering, South Essex (TQ5285) on 5 July 1999. This site is a further seven miles or so to the north. Mr Jones exhibited his specimen at a British Entomological and Natural History Society indoor meeting (Jones, 2001), incorrectly referring to the two west Kent specimens as having both been taken during the 1990s.

Whatever the original source, the occurrence of three specimens ranging over a period of some 40 years within such a relatively close geographical area, is perhaps suggestive that the species may be somewhat more than a foreign import and may be successfully established locally.

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Lathrobium fennicum Renkonen (Staphylinidae) new to Norfolk, with characters to separate females of the larger *Lathrobium* species

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While visiting Hickling Broad, Norfolk (vice-county 27), TG4221, on 2 August 2001, a single female *Lathrobium* species was collected from the base of wet vegetation at the edge of a small pool close to one of the observation hides, not far from the Visitor's Centre. It was later identified as *Lathrobium fennicum* Renkonen, a new record for Norfolk and a considerable extension in range of this species' previously known UK sites in the Isles of Scilly (MacKechnie-Jarvis, 1968) and East Sussex (Hodge, 1992).

Males of *L. fennicum* are readily separable from those of its close ally, *L. quadratum* (Paykull), using the characters of the male genitalia as illustrated, for example, by MacKechnie-Jarvis (1968) and Owen (1994) among others. Females have generally been considered more difficult to identify and Owen (1994) discussed variation in the external key characters, concluding that females should be distinguished by the shape of the eighth sternite. However, I have found that the dorsal surface of the apical (genital) segment in females provides an unequivocal character to separate *L. fennicum* and *L. quadratum*.

In conversations with various coleopterists, I have become aware that almost all experienced difficulty in separating females of the larger, common *Lathrobium* species and even Lohse (1964) noted (p. 150, couplet 5) that only the males can be identified with certainty. Newbery (1912) had also remarked on the difficulty of separating the larger females, and although he noted the female specific characters for *L. fulvipenne* (Gravenhorst) and *L. brunripes* (Fabricius), he failed to notice the differences between the other species. Unfortunately, Joy (1932) did not use Newbery's female characters in his key and, nowadays, Newbery's comments have been almost completely forgotten. In fact, the females of most of the larger species are readily separable, and therefore notes are provided on their separation as well in this paper, and not just the points of difference between *L. fennicum* and *L. quadratum*.

The last visible abdominal segment, the genital segment in Staphylinidae, has a more complex external structure than preceding segments. The staphylinid usage of, for example, Lohse (1964, Tafel 1) and Smetana (1988), among others, is followed here in regarding the genital segment to be composed of elements from both the ninth and tenth somites, such that the genital or anal tergite of some authors is

seen to represent the tenth, and not the ninth, tergite. The ninth tergite is either entire, or, more commonly, divided dorsally and developed laterally.

In both *L. fennicum* and *L. quadratum*, as also in *L. terminatum* Gravenhorst and *L. brunripes*, the ninth tergite in females is divided to the base along the dorsal midline (as Fig. 1), whereas in the other *Lathrobium* species discussed in detail below, the ninth tergite is only excised dorsally (as Figs 6-8) and not completely divided. The tenth tergite, an approximately oval apical sclerite, provides the species specific characters. In *L. quadratum*, the tergite is almost flat, or at most very weakly convex, but is bounded laterally by a submarginal ridge (Fig. 2). In *L. fennicum*, the tergite is strongly convex and sharply keeled apically (Fig. 3), thus shaped like the upturned bow of a boat. This character can usually be observed readily without the need for dissection unless the abdomen has telescoped on drying. *Lathrobium terminatum* is similar to both *L. fennicum* and *L. quadratum*, possessing similarly shaped eighth tergite and sternite and a head with a narrow neck. It is usually separable by its pale yellow outer elytral apices, but specimens with dark elytra can cause confusion. In female *L. terminatum*, the tenth tergite resembles that of *L. quadratum*, but is weakly convex and lacks the obvious submarginal ridge (Fig. 4), although a very fine lateral bead is present. In all three of the above species, the lateral processes of the female ninth tergite extend considerably beyond the apex of the tenth tergite (Fig. 1).

Lathrobium brunripes, with its broad neck cannot be confused with any of the above narrow-necked species, but its female genital segment (Fig. 5) is similar in form to the above. However, the tenth tergite is much larger, more elongate, extending back almost as far as the lateral processes of the ninth tergite, and, as noted by Newbery (1912), is longitudinally ridged. This structure immediately separates it from all the other large, broad-necked *Lathrobium* species.

The females of the large species presenting most identification problems are those four keying out from couplet 18(17) in Joy's (1932) *Lathrobium* Key 2. External body characters are subtle and not helped by variation in colour, or, especially in *L. fulvipenne*, variation in the relative length and width of the elytra. In the females of *L. fulvipenne*, the eighth sternite is broadly rounded apically, whereas in the remaining three species, it is truncated apically. The ninth tergite in female *L. fulvipenne* is excised dorsally for about the apical two-fifths of its length and the tenth tergite *in situ* appears to be almost as broad as long (Fig. 6). Both sexes possess a fringe of very fine setae along the hind margin of the elytra (Lohse, 1964), a character that appears to be reliable, if difficult to observe without good lighting and magnification, although a row of microscopic setae can just be detected in the remaining species at x70. In females of *L. geminum* Kraatz and *L. ripicola* Czwalina, the ninth tergite is excised dorsally for almost the apical half (measured along the midline), and the tenth tergite *in situ* appears to be about 1.75 times as long as broad (Fig. 7). The separation of females of these two species is more difficult and, apart from the colour differences used by Joy (1932) and others, the third antennal segment in *L. ripicola* appears proportionally longer and the succeeding segments shorter than

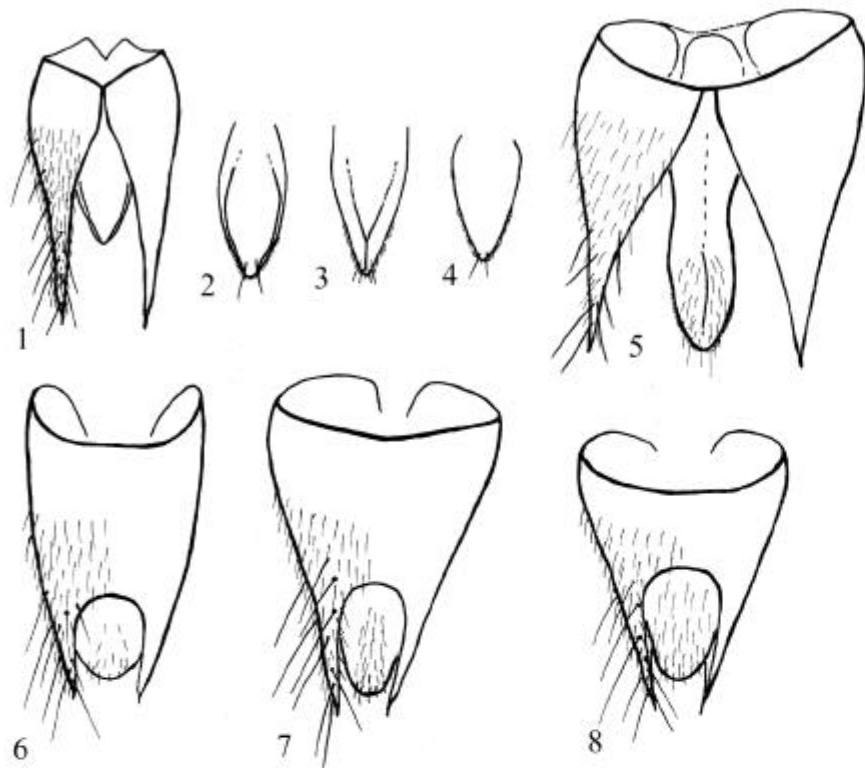
in *L. geminum*. The extent of the fine beading along the apical margin of the elytra might separate the two if found to be reliable. Examination of a small number of individuals of both sexes suggests that when viewed from directly above the body, the marginal bead extends into the inner half of the elytral hind margin in *L. ripicola*, whereas it is restricted to the outer half in *L. geminum*. Finally, in females of *L. elongatum* (Linnaeus), the ninth tergite is much more broadly and deeply excised, well into its basal half (Fig. 8) and the tenth tergite is proportionally larger and about 1.5 times as long as broad.

The female secondary sexual characters for the larger species discussed above can be summarised in the following key.

Key to females of the larger British *Lathrobium* species

1. Genital segment with ninth tergite divided dorsally (Figs. 1, 5).....2
- Genital segment with ninth tergite excised, but not divided dorsally (Figs 6-8).....4
2. Tenth tergite relatively small, much shorter than ninth tergite (as Fig. 1)....3
- Tenth tergite relatively large, almost as long as ninth tergite, longitudinally ridged (Fig. 5).....*brunripes* (Fabricius)
3. Tenth tergite almost flat, ridged laterally (Fig. 2).....*quadratum* (Paykull)
- Tenth tergite strongly convex, boat-shaped (Fig. 3).....*fennicum* Renkonen
- Tenth tergite weakly convex, not ridged laterally (Fig. 4).....
.....*terminatum* Gravenhorst
4. Ninth tergite with apical excision relatively small (Fig. 6); eighth sternite broadly rounded apically.....*fulvipenne* (Gravenhorst)
- Ninth tergite with apical excision larger (Figs 7, 8); eighth sternite truncated apically.....5
5. Ninth tergite with apical excision moderate (Fig. 7).....
.....*geminum* Kraatz and *ripicola* Czwalina (see comparative characters in text above)
- Ninth tergite with apical excision relatively large (Fig. 8).....
.....*elongatum* (Linnaeus)

A cursory examination of the tenth tergite in females of the smaller *Lathrobium* species suggests that there are subtle species-specific differences, but these species appear to key out readily without resorting to these characters. That said, *L. fovulum* Stephens is distinctive in having a sharp dorsal prominence on the tenth tergite. In most of the smaller species examined, the ninth tergite is excised (as in Figs. 6-8), but *L. pallidum* von Nordmann has a partially divided ninth tergite, more closely resembling that in Figure 1.



Figures 1-8 *Lathrobium* female genital segments, dorsal view: **1** *L. quadratum*; **2** *L. quadratum* tenth tergite; **3** *L. fennicum* tenth tergite; **4** *L. terminatum* tenth tergite; **5** *L. brunnipes* (drawn somewhat splayed); **6** *L. fulvipenne*; **7** *L. geminum*; **8** *L. elongatum*.

Although Lohse (1964) placed the narrow-necked species in the subgenus *Tetartopeus*, distinct from *Lathrobium sensu stricto*, study of the female genital segments does not entirely support this simple division. While the narrow-necked *L. fennicum*, *L. quadratum* and *L. terminatum* undoubtedly form a closely similar species group, the narrow-necked *L. angustatum* Lacordaire has a female genital segment similar to that of the majority of the broad-necked species, and the broad-necked *L. brunnipes* and *L. pallidum* have female genital segments more aligned to those of the *quadratum* group. Neck width in *Lathrobium* is a very convenient key character, but its use to define a genus-group taxon creates an artificial division which requires re-evaluation before a phylogenetically based division of the genus can be proposed.

Acknowledgement

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Review

Apionidae of the Western Palaearctic Volume 1. Introduction and Synopsis of Tribes by Mark Russell. Crocodile Press. 2001. Peterborough. ISBN 0-9541915-0-1. £80.

The first thing that strikes the reader about this book is its size - at 46 cm x 33 cm (A3) it is huge, enough to be not the coffee table book, but the coffee table itself! On opening it, one immediately discovers why. The artwork of the whole specimen illustrations is truly superb, quite the most intricately detailed and beautiful of any entomological illustration I have ever encountered, and the publishers have obviously wanted to display these highly artistic yet accurate gems in their full glory. Unfortunately, this does make for some considerable difficulty in the practical use of the book. It requires about a third of a square metre of cleared desk space beside the microscope in order to use it for what it really is: an introduction to the 13 tribes of the Apionidae of the Western Palaearctic, with keys to the tribes and all genera within those tribes.

The book is produced as a limited edition of 500 signed copies and is the first of a series aimed to describe all Palaearctic species of the genus, tribe by tribe. The author's aim is to illustrate all the species as whole body plates, which will be very welcome. At 24 pages of text it seems very short, although the very large pages means that it is the equivalent of 3 times that number of pages in a more typical sized volume. While it seems expensive for the actual page content, this reflects the quality of reproduction of the plates and also is a function of the limited size of print-run.

The book starts with an introduction and historical overview to the taxonomy of the family, a glossary of terms used and good diagrams of the structure of a typical Apionid. There follows keys to the tribes, descriptions of the individual tribes, their distribution and

food plant associations, and then keys to the genera within those tribes. Unusually, there are no line illustrations with these keys or the glossary, necessitating some to-ing and fro-ing between the keys, the glossary and the whole body illustrations in the plates. I hope that in future volumes diagnostic line illustrations will accompany keys to species. Individual representative species from each tribe (two each for Piezotrachelini and Oxystomatini) are described in detail together with their distribution and ecology and these example species form the subjects of the large, full-colour illustrations.

A brief section on philosophical problems relating to taxonomy highlights the difficulties of imposing an artificial hierarchical classificatory structure on a natural continuum, though doesn't really recognise the intended evolutionary relationships that really should be expressed in any taxonomic interpretation. The author makes no claims to be producing new taxonomic insights to the group in this work, but is bringing together existing work, illustrating it and making it available. There are occasional other philosophical and even theological thoughts of the author scattered throughout the text, in particular expressing the beauty of nature. While some may feel that these might be out of place in a scientific work on a group of insects, I find them an unusually refreshing reminder of the humanity of a scientific investigator, as is the frequent use of the first person in the text. As such, this makes for a very honest work, where the author both expresses his feelings for nature and indicates the strengths and limitations of his own work. One such note did strike me as somewhat obtuse, though. Under a separate section on 'economic importance' only the brief statement 'This aspect is beneath contempt. Man's depredations to the environment, and the mass destruction of habitat and eventually of species that this entails, will have their own peculiar payback.' An unusual statement, it would perhaps be more apt as an introductory section on conservation than on economic importance, which some species of *Protapion* undoubtedly have. Overall, however, the work is an unusual hybrid of useful science, philosophy and beautiful artwork.

A peculiarity is the 'Select Bibliography' that lists some major works on the family, but additional works are actually referred to in the text but not included in a list of references. This is an unfortunate omission with respect to a book of Vladimir Bohac's paintings which was quoted as the source of the artist's inspiration in the 1960s, but rather more serious when missing from references to the taxonomic history of the group.

There are 13 plates, each of a single representative of a tribe, all at x80 (except *Apion frumentarium* at x70). There is a separate section describing the artwork and, unusually, its limitations with respect to depiction of microsculpture, vestiture and colour and the author/artist admits to having chosen the biggest and brightest specimens to illustrate and thus introducing some bias. I understand that at least some of these plates are available as separate prints and, suitably framed, that of *Apion frumentarium* now graces the wall of our dining room.

I eagerly await future volumes.

Roger Key

Early records of *Gyrophaena poweri* Crotch (Staphylinidae) from Scotland and Wales

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Following my discovery of *Gyrophaena poweri* Crotch on the island of Rum National Nature Reserve in August 2001 and unable to trace any previous records of this species from Scotland, I published the record as new to Scotland (Welch, 2001). Some time later Geoff Hancock of the Hunterian Museum & Art Gallery at the University of Glasgow contacted me to say that, upon his recent return from the United States, he had seen my note and had checked their collections and found an earlier Scottish specimen of *G. poweri* collected by Dr. Roy Crowson. At his suggestion I not only agreed to check this specimen but also to examine the museum's entire holding of *Gyrophaena* material, with unexpected results.

The Crowson specimen had been collected on 4 October 1944 on the Bergeny Estate, Girvin, Ayrshire, on 'fungus bark', and bore the label '*Gyrophaena poweri* ♂♂ I.M. White det. 1975'. The aedeagus had been dissected out and mounted on a clear plastic strip. Although it had faded considerably in the mountant the determination was confirmed. Not unexpectedly, a number of specimens in the collections proved to be incorrectly identified or misplaced. Many of these were females that lack the distinctive external characters of the males of this genus. However, a male, lacking antennae, was found among the *G. strictula* Erichson which, when dissected, proved to be another *G. poweri* collected by R. Crowson on 8 August 1979 from a second Ayrshire locality at Dalrymple Wood, again on 'fungus bark'. A further five male specimens standing over the name *G. poweri* proved to be correctly identified. One was labelled 'Weybridge, Champion, *puncticollis* 800, *poweri* 107' and had the accession number 278 which indicated that it had come to the museum in the E. Saunders Collection. The other numbers do not relate to any of the late 19th or early 20th century published Coleoptera catalogues in my possession, and remain a mystery. I have been unable to find any previous published account of this species having been collected from this Surrey locality by G.C. Champion. The other four specimens were on one card labelled 'Llandaff, B. Tomlin, 10.98, CC (089) 4010' which came to Glasgow as part of the collections of J.J.F.X. King. The numbers 10.98 are presumed to refer to the date, October 1898. (089) is a museum accession number, and 4010 is thought to be some sort of individual catalogue or reference number used by Tomlin. There appear to be no published records of *G. poweri* from the Principality although there are accounts of its occurrence in adjoining English counties. It was recorded from Moccas Park, Herefordshire, by G.H. Ashe on 16 June 1935 (Hallett, 1951), and from Church Stretton, Shropshire, in September 1890 by W. G. Blatch (1891). The latter is not among the pre-1970 county records listed by Hyman (1994).

As the J.R. le B. Tomlin Collection is housed in the National Museum of Wales at Cardiff, Brian Levey was notified of the Glasgow discovery. He promptly checked their material and found a number of Tomlin specimens bearing the same Llandaff locality label, including the same mysterious number 4010. He dissected one of the males and confirmed it as *G. poweri* but suggested I might like to check his determination, together with those of some other specimens in their collections. The opportunity to do so did not present itself until September 2002 when I was able to confirm the identity of five male and 12 female *G. poweri* in the Tomlin Collection. Brian Levey informs me that, despite the data labels, Tomlin collected at Castell Coch, Glamorgan (ST1382), approximately 0.4 km north-west of Llandaff. Tomlin's Collection also contained specimens labelled 'Chesham' and 'White Wood, Bucks.' originating from the E.G. Elliman and B.J. MacNulty Collections. Buckinghamshire is another county not listed for *G. poweri* by Hyman (*supra cit.*). A male from a series of eight MacNulty specimens collected at Ashted, Surrey, on 1 October, 1943 standing over the name *G. poweri* was dissected and proved to be *G. hanseni* Strand. This is an early record for this species, which was not recognised as British until 1966 (Welch, 1969). Lack of time precluded close examination of the rest of this series but I have little doubt that they will all prove to be this same species.

Acknowledgements

I am indebted to both Geoff Hancock of the Hunterian Museum & Art Gallery, Glasgow, and Brian Levey of the National Museum of Wales, Cardiff, for making specimens in their care freely available to me for study.

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Some recent records of *Hypera meles* (Fabricius) (Curculionidae) in southern England

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Allen (1972) stated that this weevil is 'always very scarce with us' and summarises the few published records, all of which are very old. The species is listed in the insect Red Data Book (Shirt, 1987) as RDB3 but this status was revised to Notable A by Hyman (1992), who recorded it from 12 vice counties (six for the period 1970 onwards). Buckinghamshire, Hertfordshire, North Essex, South Devon, West Kent and West Sussex can now be added to this list. Other coleopterists must have noticed that the species is increasing in frequency because Morris (2002) states 'now regarded as locally not uncommon'. I have recorded *H. meles* (Fabricius) on numerous occasions, but mainly since 1994, and almost invariably associated with red clover. My records are listed below, by vice counties in alphabetical order.

Buckinghamshire (VC 24)

Naphill Common, SU844969, 1 male swept, 18 May 1997

East Sussex (VC 14)

Ditchling Beacon, TQ331131, 1 female swept, 11 July 1996

B2116 road verge, Plumpton, TQ368131, several off red clover, 6 June 1997

Hertfordshire (VC 20)

Bayfordbury, Hertford, TL315102, 1 male swept, 7 June 2000

North Essex (VC 19)

Moor Green, TL321267, 1 male swept, 9 June 1999

Stickling Green, TL474327, 1 male swept, 29 May 2000

North Hampshire (VC 12)

Bramshill Park, SU7660, 1 male swept, 21 June 1994

Eelmoor Marsh, SU8353, 1 male swept, 23 May 1998

South Devon (VC 3)

Bolberry Down, SX6838, 1 male swept, 26 May 1999

South Hampshire (VC 11)

West Wood, Netley, SU451094, 13 July 1986, 27 April 1998 and 17 May 1998

Claylands, Bishop's Waltham, SU547180, 2 swept, 17 May 1998

The Moors, Bishop's Waltham, SU566168, 1 female swept, 6 June 1999

Hazleton Common, SU7012, 1 male, 1 female in suction sample, 29 May 2001

West Kent (VC 16)

Lullingstone Park, TQ5164, 1 female swept, 16 June 1999

West Sussex (VC 13)

Levin Down, Singleton, SU886135, 1 male in suction sample, 13 May 1996

Mill Pond Marsh, Siddlesham, SZ858973, 1 male swept, 20 June 2002

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Leptophloeus clematidis (Erichson) (Laemophloeidae) still resident in Kent

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Mindful of old records of *Leptophloeus clematidis* (Erichson) in Kent, whenever encountered I make a point of investigating *Clematis vitalba* L. The scolytid *Xylocleptes bispinus* (Duftschmid) is invariably present but its predator had eluded me (and everyone else it seems). However, beating a stunted plant at the edge of a small wood at Leybourne, West Kent (TQ6858) I discovered a flourishing colony. Numerous adults were beaten on 5 October 2001, but by 17 October only a very few were evident.

Curiosity provoked me to split open seemingly dead twigs of about 3 mm diameter wherein both prey and predator adults were present in numbers. This observation may be a contributing factor to its apparent rarity in that it may be present externally for but a very short period and, unless one is very lucky to coincide with such an appearance, the species would not be found by general beating.

According to available published records, this beetle has only been recorded in Britain from five localities, 3 of which were situated in Kent. Despite concentrated investigation by well-known field workers, as far as I am aware it has not been recorded in Kent for nearly 90 years and it was thought that it could be 'extinct' in the county. The historical overview in Britain has recently been summarised by Barclay (2002).

Unfortunately, the *Clematis* plant stands within the route of an impending by-pass and I was asked to monitor the colony and investigate its possible distribution within the general surrounds with a view to perhaps translocating the colony to a suitable alternative location. Sadly, inspection on 8 Aug. 2002 revealed no sign of beetle activity and by 4 October 2002 the *Clematis* plant had died, possibly due to the very high water table caused by a nearby stream. Neither have I been able to locate the presence of either species in the general proximity. Maybe the scolytid is not a primary cause of the shrub's demise, but both are very transient species that only breed for a relatively short time in the already decaying plant tissues.

Reference

- BARCLAY, M.V.L. 2002. *Leptophloeus clematidis* (Erichson) (Laemophloeidae) in Surrey, with notes on its biology and an overview of its occurrence in the British Isles. *The Coleopterist* **11**(1): 21-25.

Is *Malthodes brevicollis* (Paykull) (Cantharidae) a British beetle?

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Some years ago Mike Fitton drew my attention to the absence of any British material of *Malthodes brevicollis* (Paykull) in the national collections at the Natural History Museum in London and said it was his opinion that the species should not be on the British List. I have subsequently been checking all material that I can find under the name and have come to the same conclusion.

The presence of *Malthodes brevicollis* on the British list appears to have been a mistake from the very beginning and one that has - extraordinarily - persisted to the present day. Joy (1914) introduced *M. crassicornis* (Mäklin) as British and described the differences between the two species, but appears to have not checked all previous material under the name *M. brevicollis*. *M. crassicornis* appears to have been - wrongly as it turns out - treated as an addition to the British fauna, rather than a correction of identity.

Interestingly, the male specimen of *M. crassicornis* which Joy (*op. cit.*) reported as new to the British list had been collected by P. Harwood 'near Bishops Stortford, Essex' in June 1911 and had been supposed to be *M. nigellus* Kiesenwetter (this species name was subsequently corrected to *M. brevicollis*). There is a male labelled 'Takeley, June 1911', amongst Joy material in the BMNH (M.G. Fitton, pers. comm.), which is presumably the specimen in question. It is very surprising that all earlier *M. brevicollis* material were not immediately checked. Most - if not all - of those now currently available in Museum collections have been checked in recent years and have mostly proved to be *M. crassicornis*, and **none** unequivocally *M. brevicollis*, including material from Joy's own collection as well as those of Tomlin, Power and Donisthorpe. Using the key in Fowler (1890), all *M. crassicornis* would key out to *M. nigellus* (*M. brevicollis*).

Significantly, the key in Joy (1932) perpetuates the myth of *brevicollis*, only recognising *crassicornis* from Essex. In compiling the key he must have drawn on the continental literature rather than British material. Joy (1914) only refers to having comparative material available in the form of a female *M. brevicollis* 'identified by the late Herr Ganglbauer, and lent to me by Mr. Tomlin'. This specimen is still available for inspection (see below).

Fowler (1890) appears not to have been convinced that *M. brevicollis* truly occurred in Britain. He reports that '*Malthodes nigellus*, Kies. ... was first introduced somewhat doubtfully by Mr. Crotch on two female specimens, and specimens taken by Dr Power at Mickleham and Purley oaks have been referred to it, but one of those now before me does not appear to differ from *M. atomus*'. *M. atomus*, Thomson was

the name then in current usage for what we now call *M. pumilus* (Brébisson). Much of the material in collections under the name of *M. brevicollis* is actually *M. pumilus*.

In Table 1, I have listed all records for *M. brevicollis* known to me and indicate the results of recent re-examination of all available specimens.

The main difficulty now is a specimen in the Tomlin collection at the National Museum of Wales, Cardiff. This is unfortunately a female with a deformed pronotum, but Ganglbauer was sufficiently convinced that it is indeed a *M. brevicollis* that he placed his own hand-written determination label with the specimen - it is still there. The specimen is in poor condition and is not convincingly identifiable in my opinion.

With regard to the old Yorkshire record, for which no voucher material can be found, it is of relevance that *M. crassicornis* has recently been discovered in Duncombe Park, 26 May 1987, by R.J. Marsh (pers. comm.).

Acknowledgements

Thanks to A.A. Allen for re-checking his material and for helpful correspondence, Bob Marsh for the loan of the Duncombe specimen, Peter Chandler for access to the Joy and other collections held by the British Entomological and Natural History Society, Jon Cooter (Hereford Museum), Mike Fitton (Natural History Museum), Colin Johnson (Manchester University Museum), Steve Lane (Coventry Museum), Howard Mendel (Ipswich Museum) and Peter Berridge (Torquay Museum).

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Table 1. Records for *Malthodes brevicollis* and the true identity of specimens under this name in Museum collections

Details of vice county, locality, collector, date and source	Results of checking identity of voucher specimens	Determiner
VC 3: South Devon: Woodland, Dr Leach (Stephens 1830)	none located so far	
VC 11: South Hampshire: New Forest; J.E. Black (1924)	none located so far	
VC 15: East Kent: Dungeness, A.M. Masee, 1952	about 10 specimens of <i>M. pumilus</i> in the general collection of the British Entomological & Natural History Society	KNAA
VC 17: Surrey: Wandsworth Common, Mr Westwood (Stephens 1830)	none located so far	
VC 17: Surrey: Mickleham & Purley oaks, Dr Power (Fowler 1890)	Fowler (1890) suggests these are really <i>M. pumilus</i>	
VC 18: South Essex: Hainault Forest (Donisthorpe 1926)	large <i>M. pumilus</i>	A.A. Allen
VC 18: South Essex: Waltham Forest, 17.vi.1941, A.A. Allen (1942)	female <i>M. crassicornis</i>	A.A. Allen
VC 19: North Essex: Takeley, vi.1911, P. Harwood	male <i>M. crassicornis</i> in Joy collection at BMNH	M.G. Fitton
VC 19: North Essex: 'near Bishops Stortford ¹ , Essex', vi.1911, P. Harwood (Joy (1914))	probably the same specimen as the above	

¹ Although Bishop's Stortford is in Hertfordshire, it is assumed that the records actually refer to Hatfield Forest, which lies just across the county boundary in Essex, by Takeley village. The published records all give the locality as Essex rather than Hertfordshire. Hatfield Forest is a well-known locality for *Malthodes crassicornis*.

Details of vice county, locality, collector, date and source	Results of checking identity of voucher specimens	Determiner
VC 19: North Essex: Bishop's Stortford ¹ , vi.1912, J.R. le B. Tomlin	male in Power collection, BMNH, is <i>M. crassicornis</i>	A.A. Allen & M.G. Fitton
VC 19: North Essex: Bishop's Stortford, J.R. le B. Tomlin	female in Tomlin collection, National Museum of Wales, is <i>M. crassicornis</i>	KNAA
VC 19: North Essex: Bishop's Stortford, H.C. Dolman	male in BMNH collection is <i>M. crassicornis</i>	M.G. Fitton
VC 23: Oxfordshire: Blenheim Park, 26 & 27.v.1952, G.H. Ashe	male and female <i>M. crassicornis</i> with these label details in H.W. Daltry collection, Coventry Museum	KNAA
VC 25/26: Suffolk, C. Morley & C.G. Doughty	incorrectly identified material in these collections at Ipswich Museum	KNAA
VC 26: West Suffolk: Glemsford, J.R. le B. Tomlin (Fowler & Donisthorpe 1913)	material in Tomlin collection, National Museum of Wales, is mostly <i>M. pumilus</i> plus the Ganglbauer determined deformed female	KNAA
VC 27/28: Norfolk (Hyman 1992)	none located so far	
VC 36: Herefordshire: Moccas Park, vi. 1938, G.H. Ashe [record in G.H. Ashe's annotated copy of Fowler (1890) now held by J. Cooter]	series in Ashe collection at Torquay Museum all <i>M. crassicornis</i> .	KNAA
VC 36: Herefordshire: Moccas Park, 1949, R.W. Lloyd	('Now thought to be referable to <i>M. pumilus</i> ' R.C. Welch in Harding & Wall, 2000)	
VC 36: Herefordshire: Moccas Park, 31.v.1950, R.W. Lloyd	material held by Manchester Museum not <i>M. brevicollis</i>	C. Johnson
VC 36: Herefordshire: Moccas Park, 19.v.1955, H.W. Daltry	male in Coventry Museum collection, very poor condition, not identifiable with any certainty	KNAA

Details of vice county, locality, collector, date and source	Results of checking identity of voucher specimens	Determiner
VC 36: Herefordshire: Moccas Park, 6.vi.1965, A.E. Gardner	female <i>M. crassicornis</i> in Gardner collection, National Museum of Wales	KNAA
VC 40: Shropshire: Church Stretton, Mr Blatch (Fowler 1890) [mistakenly given as in Cheshire]	none located so far	
VC 62: Mid-West Yorkshire: Bolton Woods, 1916, Fordham (G.B. Walsh, in Garland (1983))	Walsh collection at Scarborough Museum checked by L. Auckland and specimen not found	
VC 96: East Inverness: Dalwhinnie, 1913, N.H. Joy	one <i>M. pumilus</i> in the Joy collection, British Entomological & Natural History Society	KNAA
no data, T.H. Edmonds, 2 examples	material held by Manchester Museum not <i>M. brevicollis</i>	C. Johnson
no data, Mr Crotch, 2 females (Fowler 1890)	'unfortunately destroyed' according to Sharp (1868)	

Stenus fossulatus Erichson (Staphylinidae) in Scotland

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Since its discovery in County Durham in 1936 (Tottenham, 1946), *Stenus fossulatus* Erichson was not found in Britain elsewhere until 1999 when it was discovered in Cumbria (Hewitt, 2000). The Cumbrian specimens were found on and immediately below steep, unstable clay slopes.

On 4 September 2002, I found two males of the species on an unstable scar of boulder clay by the Northouse Burn, Roxburghshire (VC 80, NT439072). Although the scar was quite dry at the time, it is often flushed, wet and rather unpleasantly mobile. The beetles are a bit soft but the only visible external evidence of this is the pallid appearance of the light parts of the legs and palpi. The aedeagi are incompletely sclerotised, one being almost colourless.

This find extends the species' range into Scotland. It also adds evidence that unstable clay slopes are an important habitat, if perhaps not the only one, of the beetle.

References

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Chaetocnema picipes Stephens (Chrysomelidae) in Kent

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Since the confusion with *Chaetocnema concinna* (Marsham) was explained (Booth & Owen, 1997) I have retained a sample whenever found in the hope that I might eventually find a specimen of *C. picipes* Stephens. In fact, I can report finding it to be fairly widespread in the county of Kent as the following records demonstrate.

1. Great Stonar, Sandwich	TR3358	12 April 2001
This appears to be the first published Kent record.		
2. Leybourne varying locations within	TQ6858	28 August to 4 October 2001
3. Leeds	TQ8152	7 September 2001
4. Charing	TQ9549	22 September 2001
5. Ramsgate	TR3362	12 October 2001
6. Lyminge Forest (Denge Wood)	TR1152	11 May 2002
7. MoD- Lodge Hill, Frindsbury	TQ7473	1 June 2002
8. West Hythe	TR1234	20 July 2002

Although confirmation was possible in each case by male dissection, I found that with experience, the differences in the apical antennal segment are clear, and dissection is not necessary (although perhaps always advisable).

Reference

BOOTH, R.G. & OWEN, J.A. 1997. *Chaetocnema picipes* Stephens (Chrysomelidae Alticini) in Britain. *The Coleopterist* **6**: 85-89.

Oxytelus migrator Fauvel (Staphylinidae) in Wiltshire

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I found two examples of *Oxytelus migrator* Fauvel in a dung heap at Rushmore Park, Wiltshire, grid reference ST9519 (VC 8) on 23 September 2001. This appears to be the fourth British site (Owen, 1997) for this recent immigrant. Other immigrant species in the heap included *Perigona nigriceps* (Dejean) (Carabidae) and the staphylinids *Philonthus spinipes* Sharp and *Trichiusa immigrata* Lohse.

Acknowledgements

I thank Jane Lee for encouraging my visit and Roger Booth for help with identification.

Reference

OWEN, J.O. 1997. Some uncommon beetles from Headley Warren Surrey. *Entomologist's Record and Journal of Variation* **109**: 301-307.

Subscribers' Notices

This section is for subscribers to advertise requests for information, specimens wanted for loan, or entomological items wanted or for sale. Notices will be repeated with each issue while space is available (or until withdrawn), newer ones appearing first, and may be edited for brevity.

Lily beetle larvae wanted: I am carrying out research into the distribution of larval parasitoids of the lily beetle in the UK (See Salisbury 2003 Two parasitoids of the lily beetle, *Lilioceris lili* (Scopoli) (Coleoptera: Chrysomelidae), in Britain, including the first record of *Lemophagus errabundus* Gravenhorst (Hymenoptera: Ichneumonidae) *Br J Ent Nat Hist.* In press). I would like to receive live lily beetle larvae collected between May and September. Please send a maximum of five from any site per month to Andrew Salisbury, Entomology section, Royal Horticultural Society's Garden, **FREEPOST**, Wisley, Woking, Surrey, GU23 6BR. Please include a grid reference (minimum four figures), date of collection and host species/cultivar (if known).

From Keith Lewis During the 2002 AES Exhibition I acquired about 2000 beetles, collected in 1920-1930 plus logbooks of J. Colley. I would be most grateful for any information about this collector who I think died in 1966. kc.lewis@btoopenworld.com

For sale: *Coleopterist's Bulletin*, vol. 31 (1977) to vol. 55 (2001) inclusive: £160 the set, postage extra. Trevor G. Forsythe, 5 Knob Hill, Stretton on Dunsmore, Warwickshire. CV23 9NN Tel. 02476 542688

Help wanted: I have about 20 beetles that I cannot identify; could anyone please help? If so contact Stephen Dawson, 55 Ashbourne Crescent, Sale, Cheshire M33 3LQ Tel 0161 232 7597.

Wanted: *A monograph of the beetles associated with stored products* by Hinton 1945. Please contact Andreas Herrmann, Bremervoerder Strasse 123, D - 21682 Stade, Germany. Fax: ++49 (0) 4146 / 930083, Email: herrmann@coleopterologie.de

Wanted: Coleoptera of the British Islands by Fowler (small paper), Practical Handbook of British Beetles by Joy, Beetles of the British Isles by Linssen. Please reply to Stuart Campbell, 4 The Laurels, Moreton, Wirral, Merseyside CH46 3SU. Tel 0151 6777047 (home), 0151 4736151 (office).

Montgomeryshire beetle records wanted: I am setting out as new recorder for this vice-county and would be grateful for any records, past and present. Any information on museum or private collections containing Montgomeryshire species would also be gratefully received. *Dr W. Schaefer*, Cwm-Weeg, Dolfor Newtown POWYS SY164AT. E-mail: wolfgang@dial.l.co.uk.

New British Beetles (revised edition) by Peter Hodge & Richard Jones: Work on a new and improved version of this book is progressing well, but it would enhance its usefulness still further if the sections entitled 'Errors in Joy' were more comprehensively covered. Therefore, if you know of any previously unrecorded errors in Joy, or can supply other useful information that would make his keys work more effectively, the authors would be pleased to receive details. Reply to *Peter J. Hodge* 8 Harvard Road, Ringmer, Lewes, East Sussex BN8 SHJ. Tel.: 01273 812047.

New *Stenus* recording scheme: Please send all records of *Stenus* (Staphylinidae) species to *Jonty Denton*, 2 Sandown Close, Alton, Hampshire GU34 2TG. E-mail: jontydenton@aol.com.